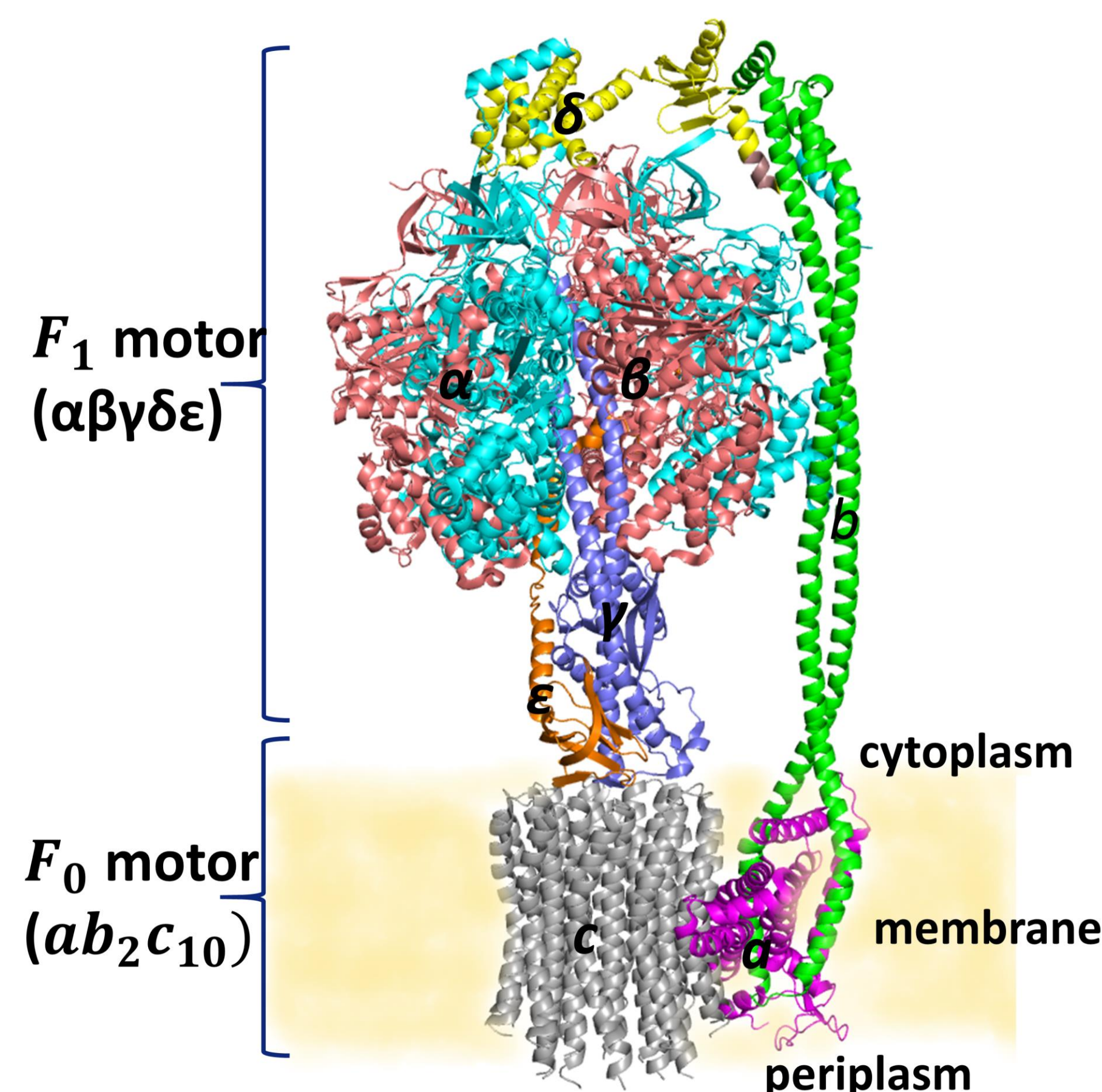


Understanding the Role of Periplasmic Loop Residues of *E. coli* ATP synthase

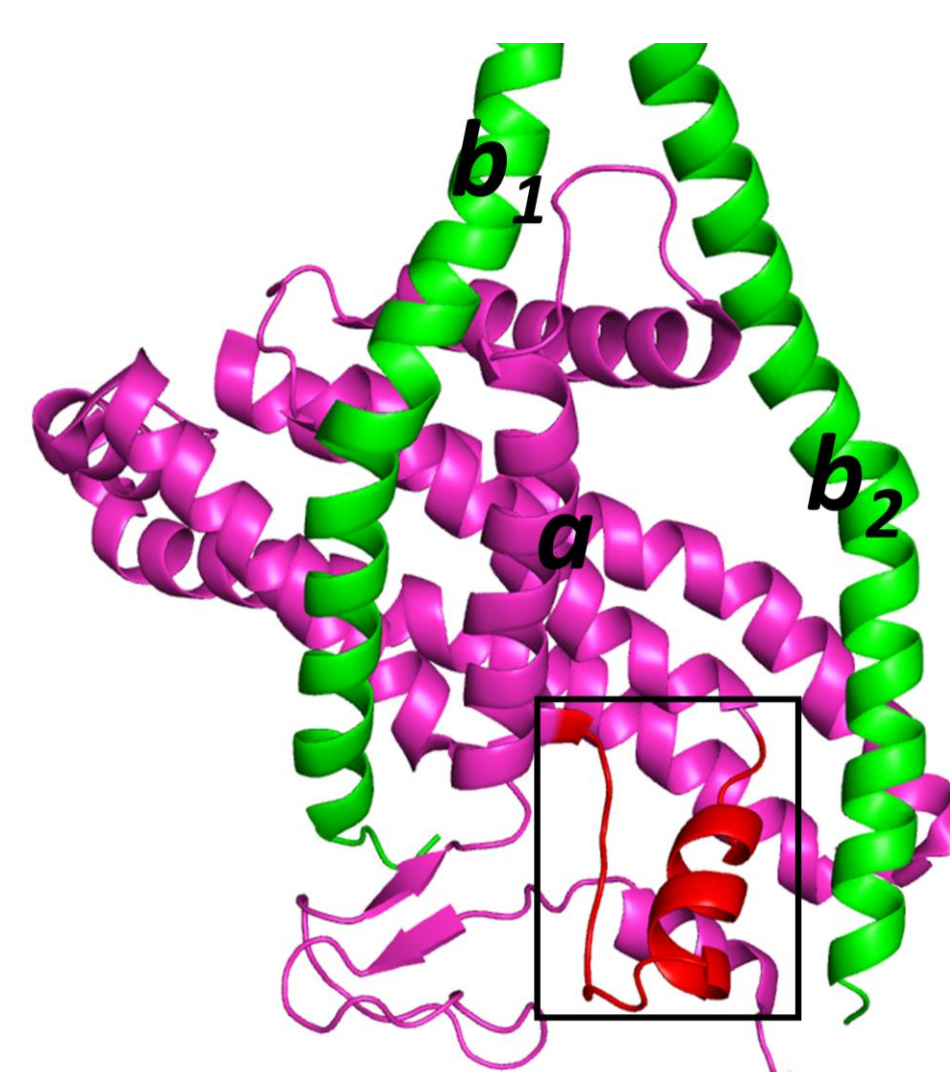
Marlene Michel, Lauren Braswell, Kingsley Basoah, Rashmi Shrestha

Department of Chemistry, Berea College, Berea, KY 40404

Background



- ATP synthase is a biological macromolecular complex that catalyzes the production of ATP.
- It consists of two unique nanomotors: a membrane bound F_0 motor and soluble F_1 motor.
- In presence of the electrochemical gradient, proton movement down the gradient rotates the F_0 motor which induces conformation change in the F_1 motor, synthesizing ATP from ADP and inorganic phosphate.
- Under certain circumstances, ATP synthase can hydrolyze ATP to transport protons against the electrochemical gradient



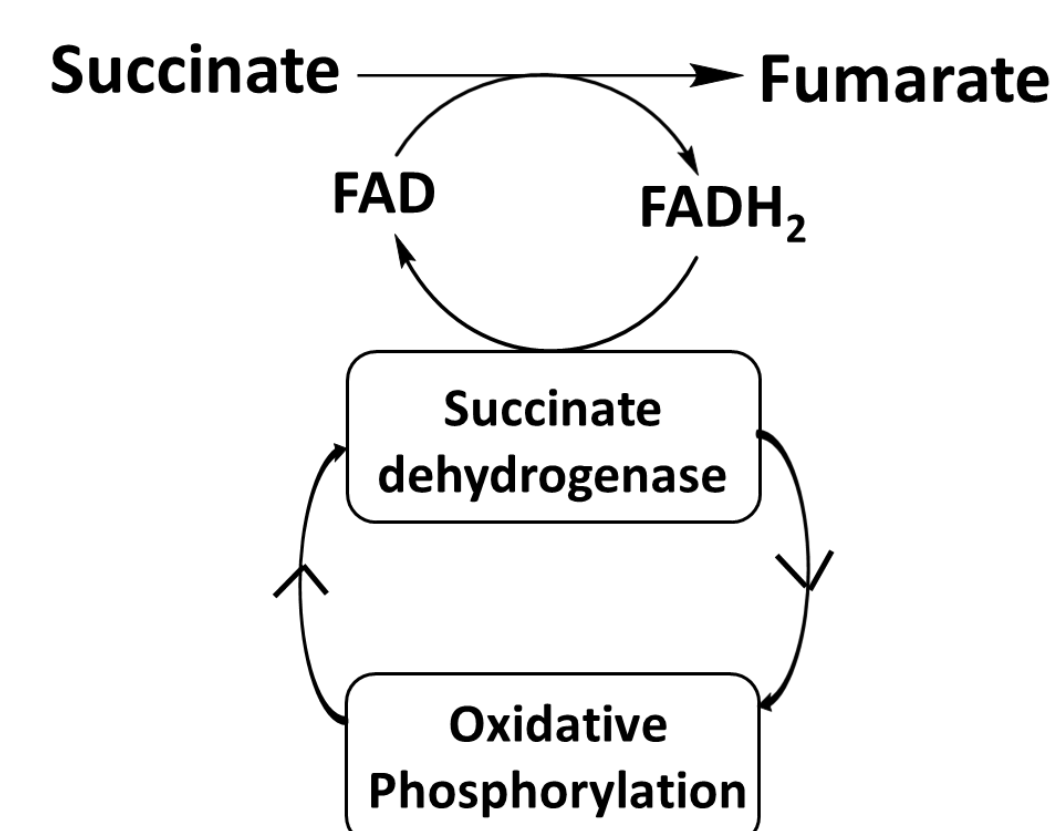
- Dysfunction of ATP synthase imbalances cellular energy, resulting in disease and even cell death.
- The increase in drug-resistant bacterial infections necessitate the identification of novel drug target sites on ATP synthase.
- Difference in sequence and composition of TM 2-3 loop of subunit a across species may help selectively target ATP synthase of different bacteria.

	TM 2-3 loop
ECOLI	LMDLLPIDLLPYIAEHVGLPALRVVPSADVNTLS
BACP3	MLGLPFS-----VHVNG-ELIWKSPATADATVTLI
MYCTU	WLAVLPVQYT---DKHGHT-TELLKSAADINVVL

Figure 1: Subunit a of bacterial ATP Synthase: A) Residues T M 2-3 loop, shown in red, suggested to interact with subunit b_2 . B) Sequence alignment of subunit a from *E. coli*, *Bacillus PS3* and *Mycobacterium tuberculosis*, highlighting the TM 2-3 loop.

Growth on Succinate

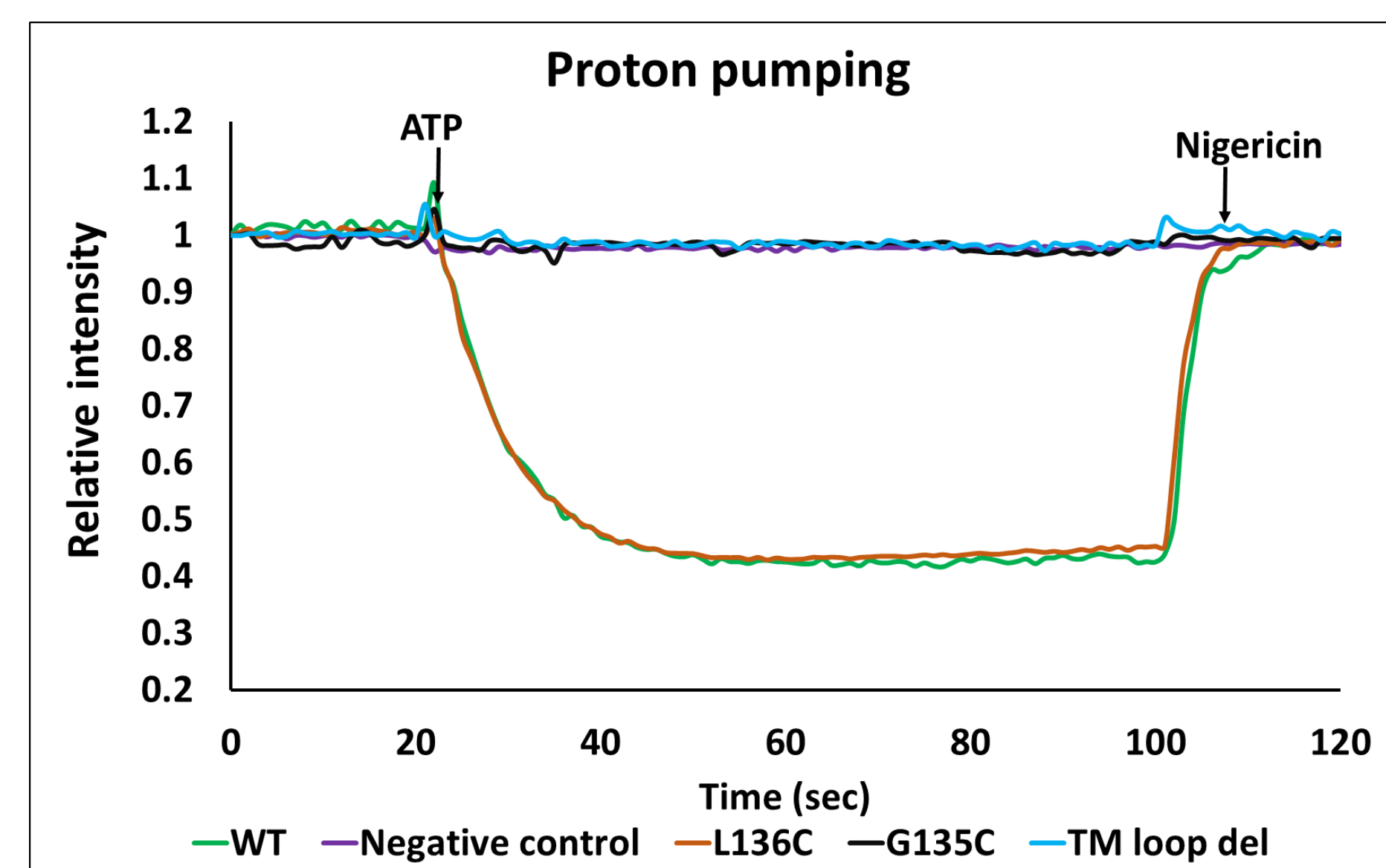
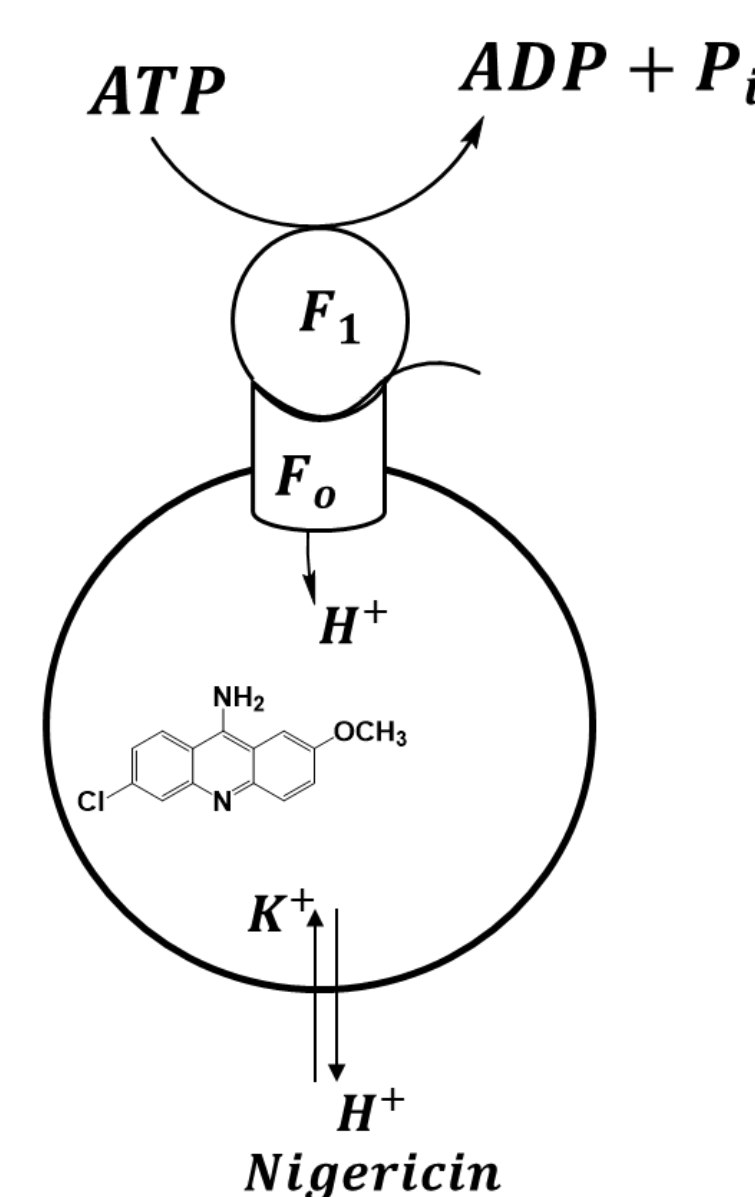
- Succinate is a nonfermentable carbon source that can be metabolized only in presence of functional ATP synthase
- Colony size is measured to determine defect in growth
- Slight defects in growth is seen for TM loop delete, G135C and L136C mutants



Cell Type	Growth	Colony Size
Wild Type	Positive	2 mm
Negative Control	Negative	N/A
TM loop del	Negative	1.5 mm
G135C	Positive	1.75mm
L136C	Positive	1.75 mm

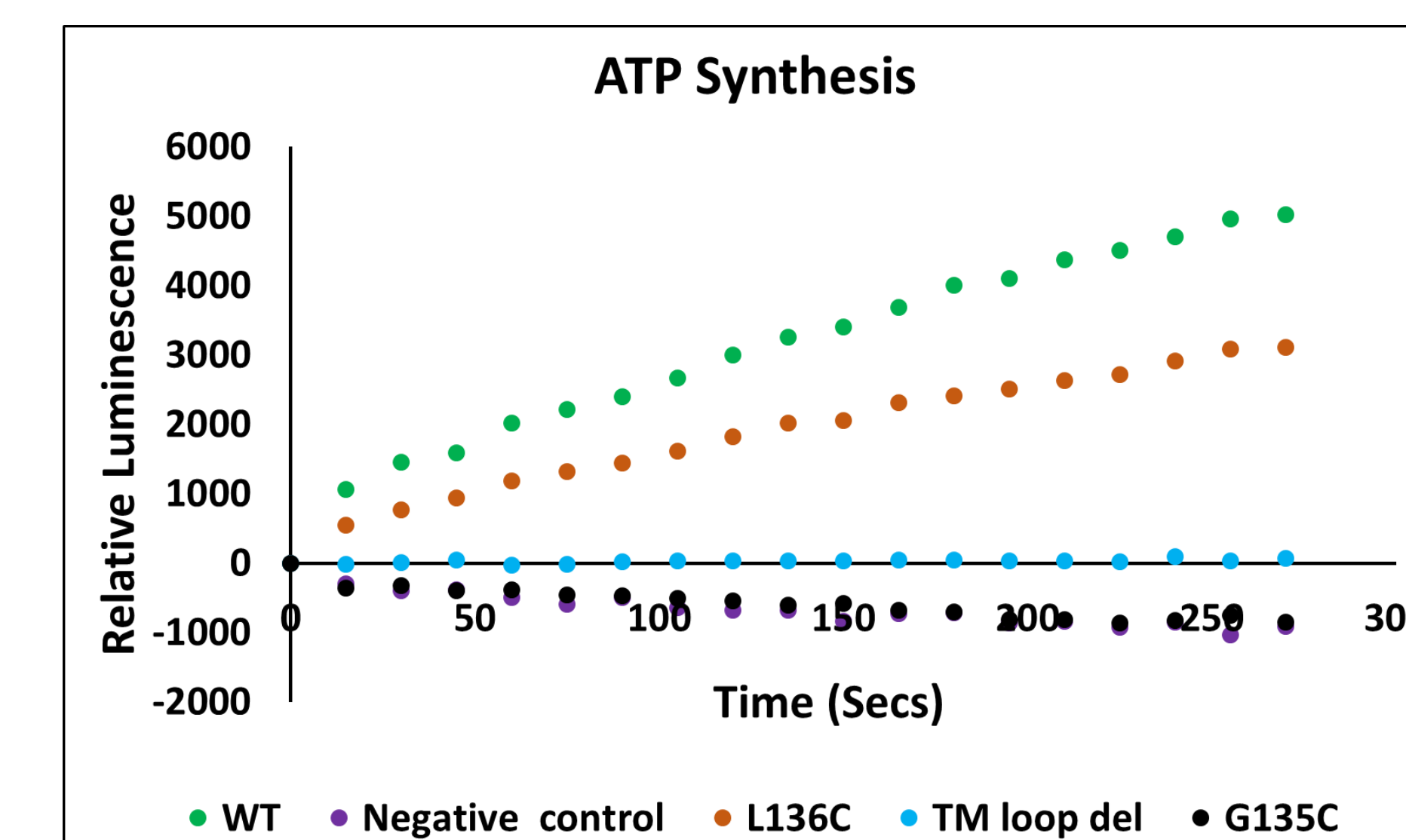
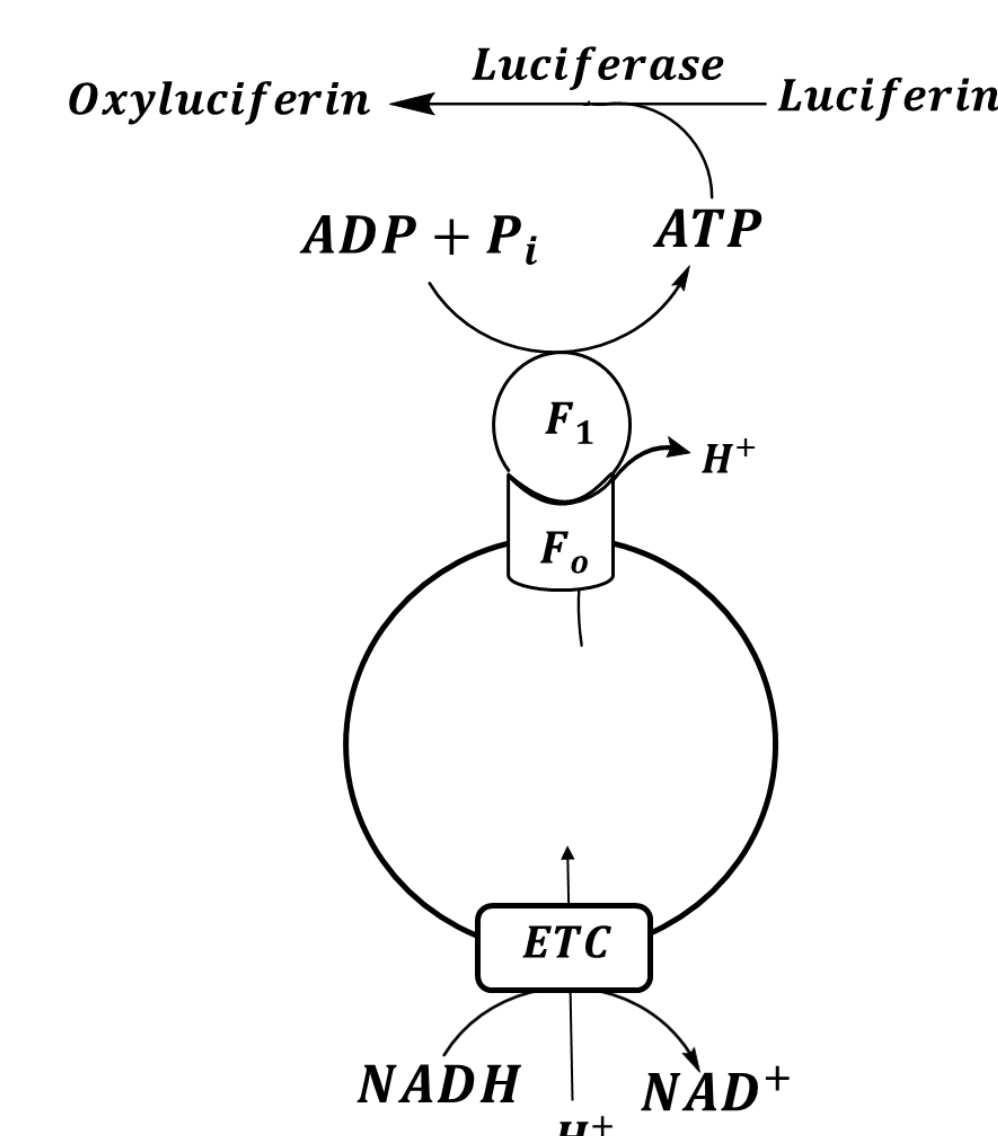
ATP-driven Proton Pumping

- Relies on the fluorescence of ACMA dye, which freely diffuses through the membrane
- Addition of ATP pumps proton into the vesicle and protonates ACMA, quenching the fluorescence
- Addition of nigericin destabilizes the gradient restoring fluorescence
- G135C and TM loop delete mutants inhibit proton pumping
- L136C is able to pump protons similarly to wild type



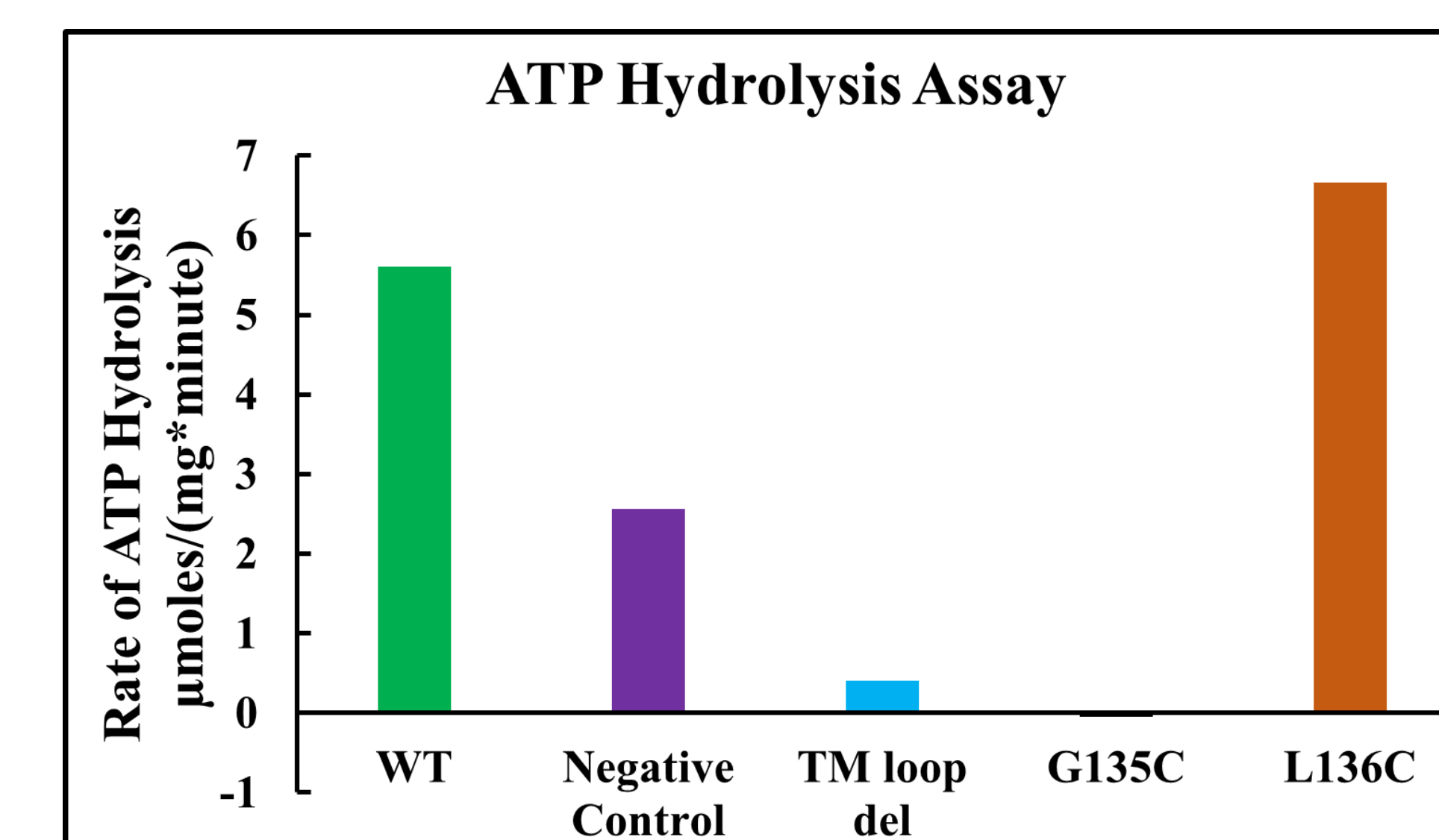
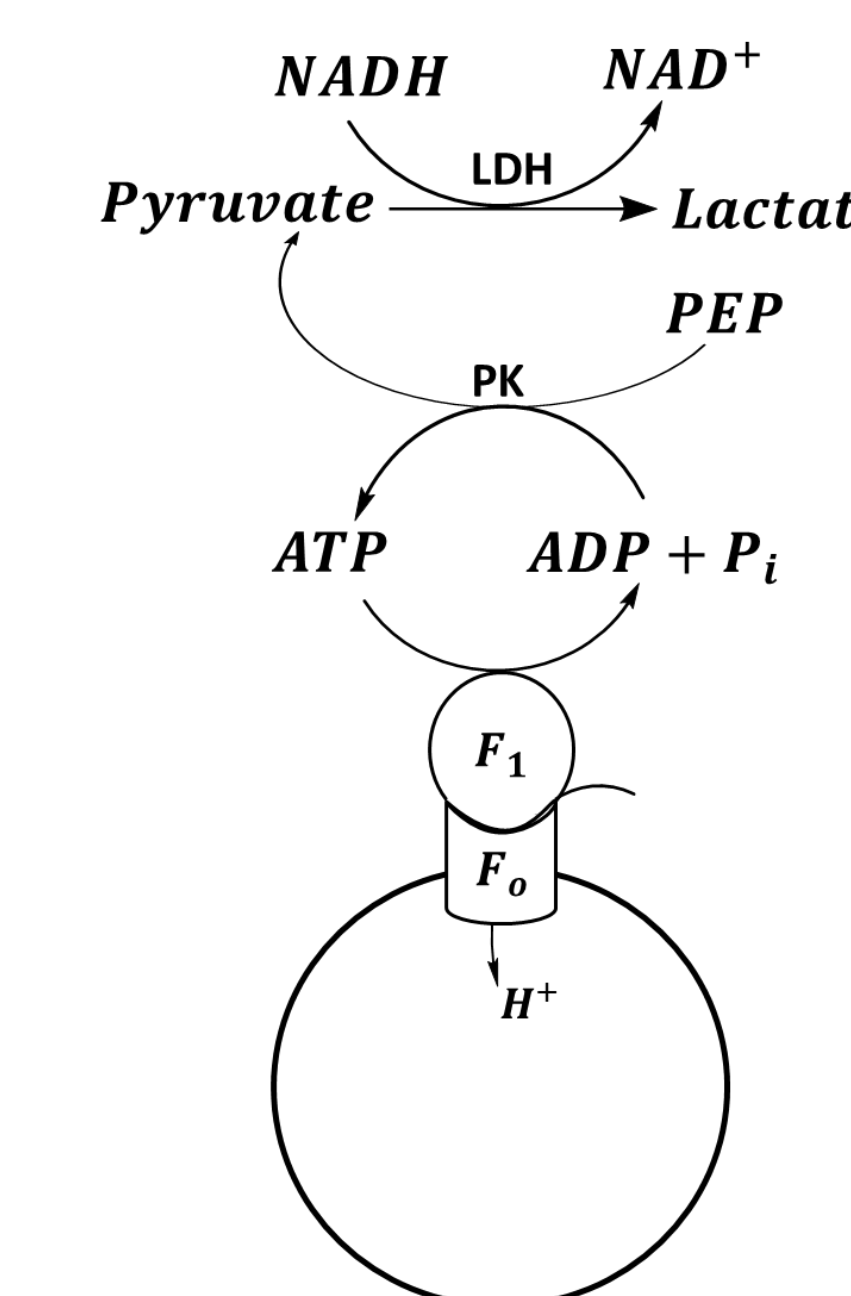
ATP Synthesis

- Concentration of ATP over time is measured using ATP-dependent luciferase/luciferin system
- NADH activates electron transport chain (ETC) and pumps proton into the vesicle driving synthesis of ATP, which is required for oxidation of luciferin to oxyluciferin.
- G135C and TM loop delete mutants show no ATP synthesis activity
- L136C mutant shows decrease in luminescence indicating slight defect in ATP synthesis



ATP Hydrolysis

- Rate of ATP hydrolysis is measured using NADH-linked ATP-regeneration system.
- ATP hydrolysis is coupled to NADH oxidation via lactate dehydrogenase and pyruvate kinase.
- G135C and TM loop delete mutants show defect in ATP hydrolysis consistent with the proton pumping assay
- L136C enhances the rate of hydrolysis.



Conclusion/ Future Directions

- TM 2-3 loop delete mutant affects all the activities of ATP synthase suggesting its importance for the function of the enzyme.
- G135C mutation suggests that glycine at position 135 seems to be an important residue. However, additional mutations will give more insight to the essential characteristics of this residue.
- L136C mutation reveals asymmetric effects on proton movement.
- The experiments with these residues will need to be replicated for validation as well as the expression levels of each mutant needs to be determined.
- We need to optimize the ATP hydrolysis assay to decrease the activity of negative control.

Acknowledgement & References

The authors acknowledge support from Undergraduate Research and Creative Projects Program, Berea College Department of Chemistry and Geology, and Dr. David Rodgers for providing access to instrumentation at University of Kentucky.

- Ishmukhametov, R. R., DeLeon-Rangel, J., Zhu, S. & Vik, S. B. Analysis of an N-terminal deletion in subunit a of the Escherichia coli ATP synthase. *J Bioenerg Biomembr* **49**, 171-181 (2017).
- Guo, H., Suzuki, T. & Rubinstein, J. L. Structure of a bacterial ATP synthase. *Elife* **8** (2019).
- Steed, P. R., Kraft, K. A. & Fillingame, R. H. Interacting cytoplasmic loops of subunits a and c of Escherichia coli F1F0 ATP synthase gate H+ transport to the cytoplasm. *Proc Natl Acad Sci U S A* **111**, 16730-16735 (2014).
- Sobti, M., Walshe, J.L., Wu, D. et al. Cryo-EM structures provide insight into how E. coli F1F0 ATP synthase accommodates symmetry mismatch. *Nat Commun* **11**, 2615 (2020).
- Angevine C.M., Herold K.A.G., Vincent O.D. & Fillingame R.H. Aqueous Access Pathways in ATP Synthase Subunit a Reactivity of Cysteine Substituted into Transmembrane Helices 1, 3 and 5. *J. Biol. Chem.* **282**, 9001-9007 (2006).